

# Insights to the formation of secondary inorganic PM<sub>2.5</sub>: Current knowledge and future needs

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**Abstract:** As a criteria pollutant, fine particulate matter (fine PM, i.e. PM<sub>2.5</sub>) adversely affects public health and environment, contributes to visibility degradation and regional haze. Atmospheric fine PM includes primary and secondary PM<sub>2.5</sub>. While the primary PM<sub>2.5</sub> is from direct emissions, the secondary PM<sub>2.5</sub> is formed in the atmosphere through photochemical reaction, condensation and other atmospheric processes. Although it is well known that ammonia (NH<sub>3</sub>) may react with acidic gas species to form secondary inorganic PM<sub>2.5</sub> (iPM<sub>2.5</sub>) as ammonium salts, limited research has been done to quantify the impacts of NH<sub>3</sub> emissions of animal feeding operations (AFOs) on the dynamics of such chemical reactions and gas-particle phases partitioning. This paper is to provide comprehensive review of existing research on AFO PM chemical speciation and on the formation of secondary iPM<sub>2.5</sub> as impacted by AFO air emissions. Research gaps and future studies in characterizing AFO PM and assessing impacts of AFO air emissions on atmospheric PM are discussed.

**Keywords:** environmental pollution, particulate matter, inorganic PM<sub>2.5</sub>, animal feeding operations, ammonia, formation of secondary PM<sub>2.5</sub>

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## 1 Introduction

Fine particulate matter (PM) refers to liquid and/or solid particles that are suspended in the air with aerodynamic equivalent diameter (AED) smaller than or equal to 2.5  $\mu\text{m}$  (i.e. PM<sub>2.5</sub>). Studies indicate that PM<sub>2.5</sub> may cause serious environmental and health problems<sup>[1-7]</sup>. Exposure to PM<sub>2.5</sub> has been linked to a variety of lung and heart diseases that may lead to cardiovascular and respiratory mortality and premature death<sup>[1-6]</sup>. It was estimated that a 10  $\mu\text{g}/\text{m}^3$  increase in 2-day averaged PM<sub>2.5</sub> mass concentration may cause a 0.74% increase in non-accidental deaths<sup>[6]</sup>. In addition to the adverse

health effects, PM<sub>2.5</sub> may also cause ambient air quality reduction and visibility impairment<sup>[8-9]</sup>. In atmosphere, particles with sizes in range of the light wavelength are more efficient to scatter and absorb the lights, causing atmospheric visibility degradation. Therefore, fine PM is a major contributor to regional haze and smog problems<sup>[8-10]</sup>.

Based upon its physical and chemical properties as well as the formation mechanism, atmospheric PM may be classified as (1) fine vs. coarse PM; (2) organic vs. inorganic PM; (3) primary vs. secondary PM. The primary PM is emitted directly to the atmosphere whereas the secondary PM is formed in the atmosphere through photochemical reaction, condensation and other atmospheric processes<sup>[10-11]</sup>. Both primary and secondary PM may be organic and/or inorganic in nature. While most primary PM is coarse PM (AED > 2.5  $\mu\text{m}$ ), secondary PM is fine in size (AED  $\leq$  2.5  $\mu\text{m}$ , i.e. PM<sub>2.5</sub>).

On average, inorganic PM<sub>2.5</sub> (iPM<sub>2.5</sub>) makes up about half of total atmospheric PM<sub>2.5</sub> in the United States<sup>[12-17]</sup>,

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and 20%-30% of total  $PM_{2.5}$  mass in urban environments in China<sup>[18-21]</sup>. Very little  $iPM_{2.5}$  is primary<sup>[22]</sup> and most of atmospheric  $iPM_{2.5}$  is secondary, formed through acid/base neutralization processes, in which base  $NH_3$  reacts with acidic gases, e.g.,  $SO_2$  and/or nitrogen oxides ( $NO_x$ ) to form aerosols in  $(NH_4)_2SO_4$  or  $NH_4HSO_4$  or  $NH_4NO_3$ . In these ammonium salts formation processes, partitioning of gas-particle phases ( $NH_3-NH_4^+$ ) is highly dependent on the temperature, relative humidity (RH), and availabilities of the precursor gases (i.e.  $NH_3$ ,  $SO_x$ ,  $NO_x$ )<sup>[11,22]</sup>. Therefore, control of atmospheric  $PM_{2.5}$  requires systematical understanding of  $iPM_{2.5}$  formation and the role of its precursor gas emissions.

As the most abundant base gas in atmosphere,  $NH_3$  plays a very critical role in the formation of secondary  $iPM_{2.5}$ . Although it remains to be further validated, Pinder and Adams<sup>[22]</sup> suggested that control of  $NH_3$  emissions may be used as a cost-effective supplementary strategy to reduce atmospheric  $iPM_{2.5}$ . Sources of atmospheric  $NH_3$  include animal feeding operations (AFOs) (e.g. livestock, etc.), fertilizer application, fuel combustion, industrial processes, and others. As illustrated in Figure 1, AFOs is the largest source, responsible for approximate 80% of total  $NH_3$  emissions in the U.S. Impact of AFO  $NH_3$  emissions on the formation of atmospheric  $iPM_{2.5}$  cannot be overlooked.

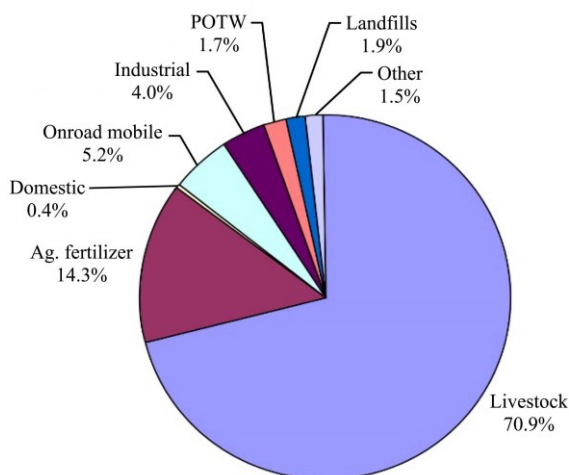


Figure 1 Contributions of various  $NH_3$  emission sources to the U.S. National Emission Inventory (NEI)<sup>[23]</sup>  
(POTW: publicly owned treatment works)

Animal feeding operation facilities may emit large amount of gaseous pollutants ( $NH_3$ ,  $H_2S$ , VOCs, and odor) and PM (TSP,  $PM_{10}$ , and  $PM_{2.5}$ )<sup>[24-26]</sup>, they have been

considered significant sources of atmospheric air pollutants<sup>[27]</sup>. In particular, PM emitted from AFO facilities may be classified as primary and secondary PM based upon its formation mechanism. It has been reported that PM in AFO houses is mainly primary, coarse and organic in nature<sup>[28-34]</sup>. In addition to direct emissions of PM, gaseous pollutants emitted from AFO facilities may be important precursors of secondary PM (organic and inorganic), which may represent a significant fraction of  $PM_{2.5}$ <sup>[31,32,35-38]</sup>.

Although the contributions of AFO air emissions to atmospheric  $PM_{2.5}$  have not been experimentally quantified and impact of AFO precursor gas emissions on the formation of secondary  $PM_{2.5}$  is not well understood, AFOs have been perceived to be important contributors to atmospheric  $PM_{2.5}$ <sup>[7]</sup>. This review paper aims to provide an overview of existing research on  $PM_{2.5}$  chemical speciation and the formation of secondary  $iPM_{2.5}$  as impacted by the  $NH_3$  emissions from AFOs, the most important contributor to the atmospheric  $NH_3$ . Research gaps and future studies in characterizing AFO PM and assessing impacts of AFO air emissions on atmospheric PM are discussed.

## 2 $PM_{2.5}$ chemical compositions

Fundamental understanding of  $PM_{2.5}$  source apportionment and formation of secondary  $PM_{2.5}$  requires knowledge of  $PM_{2.5}$  chemical compositions. In atmosphere, different areas and regions have different source types, climatology and geography; therefore, different concentrations and chemical/physical characteristics of ambient  $PM_{2.5}$ . Consequently, wide spatial and temporal variations in  $PM_{2.5}$  concentrations and characteristics exist. In chemical characterization,  $PM_{2.5}$  mainly consists of inorganic constituents (e.g.  $NH_4^+$ ,  $SO_4^{2-}$ ,  $NO_3^-$ ,  $Na^+$ ,  $K^+$ ,  $Cl^-$ , etc.), carbon species (i.e. organic carbon (OC), element carbon (EC)), and trace elements (geological materials e.g., Al, Si, Ca, Ti, Fe, etc.). The ionic species of  $PM_{2.5}$  link to various chemical reactions that lead to formation of secondary inorganic  $PM_{2.5}$ <sup>[39]</sup>.

### 2.1 Ambient $PM_{2.5}$ chemical speciation monitoring networks in the U.S.

Chemical speciation provides characterization of ions,

carbon species, and trace elements of PM. The approaches used in PM<sub>2.5</sub> chemical speciation include both field sampling and laboratory analysis. The full set of ambient PM<sub>2.5</sub> chemical speciation analysis includes examination of ions (NH<sub>4</sub><sup>+</sup>, SO<sub>4</sub><sup>2-</sup>, NO<sub>3</sub><sup>-</sup>, Na<sup>+</sup>, K<sup>+</sup>, Cl<sup>-</sup>), organic and elemental carbons (OC/EC), and trace elements (Na-Pb)<sup>[40,41]</sup>. In the U.S., three major chemical speciation networks provide complementary data on PM<sub>2.5</sub> chemical compositions to support PM source identification, implementation plan development, and health effects research. These three networks include the Interagency Monitoring of Protected Visual Environments (IMPROVE), the Clean Air Status and Trends Network (CASTNET), and the National Chemical Speciation Network (CSN). The IMPROVE network was established in 1988 to monitor mass concentrations and chemical compositions of PM<sub>2.5</sub> in the National Parks and other visibility protected rural areas for assessment of visibility degradation<sup>[42]</sup>. Data from IMPROVE have been used to calculate Haze Index in support of the USEPA's Regional Haze Rule<sup>[8]</sup>. The CASTNET was established in 1991 to monitor concentrations and depositions of sulfur and nitrogen species (gas and particulate phases) as well as ozone concentrations<sup>[43]</sup>. Data from the CAETNET have been used to assess long term trends in atmospheric sulfur and nitrogen pollutant concentrations and their depositions and ecological effects in response to changes in air emissions. The CSN began in 1999 to monitor mass and chemical compositions of PM<sub>2.5</sub> at representative sites in urban areas in the U.S.<sup>[44]</sup>. PM<sub>2.5</sub> analytes of the CSN are similar to those of the IMPROVE, consisting of an array of ions, carbon species, and trace elements. Data from 54 National Air Monitoring Stations (NAMS) of the CSN have been used for assessment of trends and data from 250 of State and Local Air Monitoring Stations (SLAMS) of the CSN provide information for developing effective State Implementation Plans (SIPs). Due to the different data use objectives, the target species of interest and analytical methods are different at these three speciation programs (networks). Table 1 provides a brief summary of the measurement parameters by the three networks<sup>[45]</sup>.

Although these monitoring networks offer high

quality PM chemical speciation datasets for air quality and public health studies, there is a lack of representation of agricultural operation areas to assess ambient iPM<sub>2.5</sub> as impacted by AFO NH<sub>3</sub> emissions, which account for majority of the nation's NH<sub>3</sub> emissions.

**Table 1 Summary of the parameters measured by the three chemical speciation networks**

	IMPROVE	CASTNET <sup>a</sup>	CSN
Sampling inlet heads	PM <sub>10</sub> head + SCC <sup>b</sup> PM <sub>2.5</sub> head	Open faced inlet	PM <sub>10</sub> head + SCC <sup>b</sup> PM <sub>2.5</sub> head
Sampling flow rate	22.8 LPM	1.5 LPM	6.7 LPM
Teflon Filter	PM <sub>2.5</sub> mass, trace elements <sup>c</sup>	Mass of non-size- selective PM, NO <sub>3</sub> <sup>-</sup> , SO <sub>4</sub> <sup>2-</sup> , NH <sub>4</sub> <sup>+</sup> , Na <sup>+</sup> , K <sup>+</sup> , Mg <sup>2+</sup> , Cl <sup>-</sup>	PM <sub>2.5</sub> mass, trace elements <sup>c</sup>
Nylon Filter	PM <sub>2.5</sub> ions: SO <sub>4</sub> <sup>2-</sup> , NO <sub>3</sub> <sup>-</sup> , Na <sup>+</sup> , K <sup>+</sup> , Cl <sup>-</sup> , NH <sub>4</sub> <sup>+</sup> (calculated)	HNO <sub>3</sub> , SO <sub>2</sub>	PM <sub>2.5</sub> ions: SO <sub>4</sub> <sup>2-</sup> , NO <sub>3</sub> <sup>-</sup> , Na <sup>+</sup> , K <sup>+</sup> , Cl <sup>-</sup> , NH <sub>4</sub> <sup>+</sup>
Quartz Filter	PM <sub>2.5</sub> OC/EC <sup>d</sup>	--	PM <sub>2.5</sub> OC/EC <sup>d</sup>

Note: <sup>a</sup> CASTNET also includes measurements of gas pollutants (e.g. NO/NO<sub>y</sub>, SO<sub>2</sub>, CO, O<sub>3</sub>), and meteorological parameters<sup>[46]</sup>.

<sup>b</sup> SCC: sharp cut cyclone.

<sup>c</sup> Up to 48 elements (sodium through lead) are determined by Energy dispersive x-ray fluorescence (XRF).

<sup>d</sup> OC/EC operationally defined by a thermal-optical analysis method.

## 2.2 Chemical compositions of AFO PM

Although knowledge about PM source profile and chemical compositions is essential for source identification and development of source-specific PM mitigation techniques, study of AFO PM source apportionment and chemical speciation is very much limited. Aarnink et al.<sup>[47]</sup> reported that PM in livestock buildings is mainly composed of primary PM with organic matter in nature. Chemical compositions of PM in animal houses varied with animal species, housing type, waste management systems (e.g. dry or wet systems, with or without bedding materials, house cleaning methods, etc.). According to Aarnink et al.<sup>[47]</sup>, every kilogram inhalable PM in swine houses contained 920 g of dry matter, 149.5 g ash, 67 g N, 14.7 g P, 27.8 g K, 7.8 g Cl and 8.2 g Na, respectively; every kilogram inhalable PM in broiler houses contained 911 g of dry matter, 97.4 g ash, 169 g N, 6.4 g P, 40.3 g K, 4.2 g Cl and 3.2 g Na, respectively (illustrated in Figure 2). Cambra-Lopez et al.<sup>[29,30,48]</sup> used particle morphological and chemical speciation data to identify and quantify the contributions of various sources to primary PM<sub>2.5</sub> and PM<sub>coarse</sub> (i.e., PM<sub>10-2.5</sub>) in poultry and swine production houses. They

discovered that feather and manure are two major sources of PM in poultry houses, skin and manure are major sources of PM in swine houses (Table 2). Contribution from feed to both fine and coarse PM was insignificant.

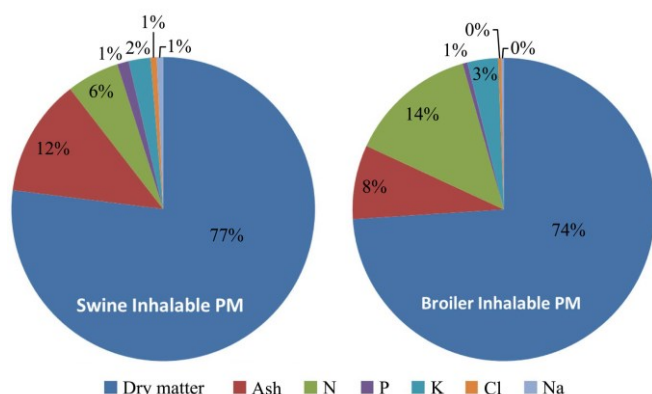


Figure 2 Compositions of inhalable PM in swine and broiler production houses<sup>[47]</sup>

**Table 2** Relative percentage contributions (by mass) of source type to PM<sub>2.5</sub> and PM<sub>10-2.5</sub> in 21 animal production houses

House type	Source type <sup>c</sup>	PM <sub>2.5</sub>	PM <sub>10-2.5</sub>
Poultry <sup>a</sup>	feather	17%-68%	4%-49%
	manure	6%-77%	31%-96%
Swine <sup>b</sup>	skin	0-79%	0-71%
	manure	14%-95%	23%-92%

Note: <sup>a</sup> Poultry houses: 3 houses of broiler on bedding, 3 house of laying hens on floor, 3 houses of laying hens in aviary and 3 houses of turkey on bedding.

<sup>b</sup> Swine houses: 3 houses of piglets on slatted floor, 3 houses of growing-finishing pigs on partially slatted floor, 3 houses of dry and pregnant sows on slatted floor.

<sup>c</sup> Elements of N, Na, Mg, Al, Si, P, S, Cl, K and Ca presented in all sources and their relative concentrations were different among sources.

In a study of trace elements and soluble ions in PM<sub>2.5</sub> and PM<sub>10</sub> emitted from 12 swine houses and 6 poultry houses in Midwest of U.S., Yang et al.<sup>[49]</sup> reported that 19 trace elements and 4 inorganic ions (Cl<sup>-</sup>, NO<sub>3</sub><sup>-</sup>, SO<sub>4</sub><sup>2-</sup> and NH<sub>4</sub><sup>+</sup>) were identified in PM samples taken from the animal production houses. Concentration of NH<sub>4</sub><sup>+</sup> in PM samples was less than 1% wt indicating insignificant formation of NH<sub>4</sub><sup>+</sup> containing secondary PM<sub>2.5</sub>. In addition, significant differences in inorganic compositions were observed between PM<sub>10</sub> and PM<sub>2.5</sub> collected from swine and turkey houses; seasonal variation of inorganic composition in PM<sub>10</sub> was insignificant, but the seasonal variation of PM<sub>2.5</sub> inorganic composition was significant<sup>[49]</sup>.

Elemental concentrations of PM<sub>10</sub> and PM<sub>2.5</sub> were first investigated at a commercial dairy farm in Californian of U.S.<sup>[50]</sup> It was discovered that Si, Al, Mg,

Fe, Ca, K, S and Cl accounted for 95.6% and 97.9% of the total elemental concentrations in PM<sub>2.5</sub> and PM<sub>10</sub>, respectively. Significant differences in percentages of each element under dry and wet conditions were also observed. Different emission source types (e.g. drylot corrals, upwind freeway, etc.) and strength under dry and wet conditions were responsible for the variations of elemental concentrations and abundances of elemental compositions.

Chemical compositions of PM in animal houses (at source) and in the vicinity of the houses (in ambient) have seldom been studied simultaneously. In an investigation of PM<sub>2.5</sub> chemical speciation in an egg production house and at five ambient locations surrounding the production houses, Li et al.<sup>[32,36-38,51,52]</sup> reported that in the production house, NH<sub>4</sub><sup>+</sup>, SO<sub>4</sub><sup>2-</sup> and NO<sub>3</sub><sup>-</sup> accounted for 10% of the PM<sub>2.5</sub> mass, but at ambient locations, NH<sub>4</sub><sup>+</sup>, SO<sub>4</sub><sup>2-</sup> and NO<sub>3</sub><sup>-</sup> accounted for 36%-41% of the PM<sub>2.5</sub> mass. In house PM<sub>2.5</sub> had much higher concentrations of Ca, Cl, K, Mg, Na, P, S, Si and Zn than those of ambient PM<sub>2.5</sub> samples. All sampling locations (in-house and ambient stations) had very low concentrations of Ag, Al, As, Ba, Cd, Ce, Co, Cr, Cs, Cu, In, Ni, Pb, Rb, Sb, Se, Sn, Sr, Ti, V and Zr. Different elemental composition distribution patterns were observed in PM<sub>2.5</sub> samples taken at the in-house station and the ambient stations (Figure 3). Majority components in PM<sub>2.5</sub> at this AFO site were organic. On average, OC accounted for above 50% of PM<sub>2.5</sub> mass at in-house and ambient stations; NH<sub>4</sub><sup>+</sup>, SO<sub>4</sub><sup>2-</sup> and NO<sub>3</sub><sup>-</sup> accounted for about 40% of the total PM<sub>2.5</sub> mass in ambient locations and for only 12% of the total PM<sub>2.5</sub> mass in house.

### 3 Formation of secondary inorganic PM<sub>2.5</sub>

#### 3.1 Ammonia as a precursor to secondary inorganic PM<sub>2.5</sub>

Major inorganic constituents of iPM<sub>2.5</sub> are NH<sub>4</sub>NO<sub>3</sub>, (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> and NH<sub>4</sub>Cl<sup>[12-22]</sup>. These constituents were formed through chemical reactions of NH<sub>3</sub> with acidic gases (e.g. SO<sub>2</sub> and NO<sub>x</sub>)<sup>[11,53-55]</sup>. Figure 4a shows a depiction of the iPM<sub>2.5</sub> formation potential in response to the major precursor gases, and Figure 4b illustrates a simplified version of the chemical reaction processes to the formation of iPM<sub>2.5</sub>.

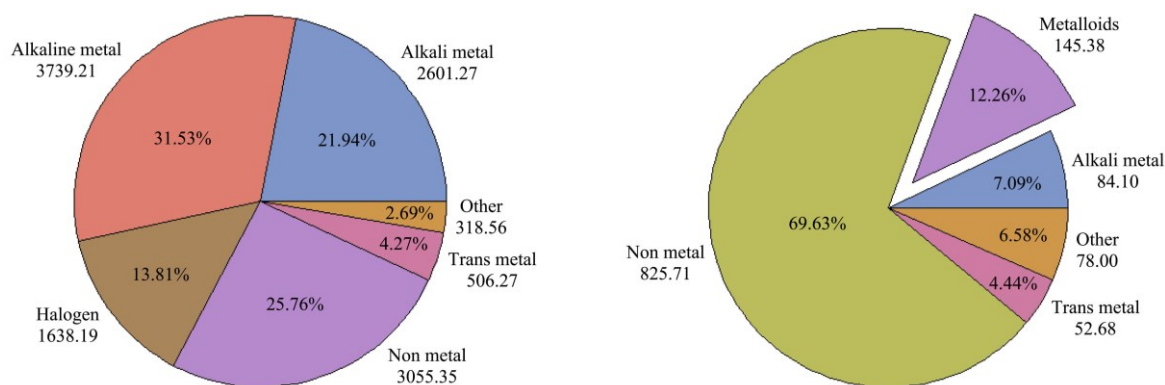


Figure 3 Elemental compositions of PM<sub>2.5</sub> in a production house (left) and at an ambient location in the vicinity of the egg production farm (right)<sup>[38]</sup>

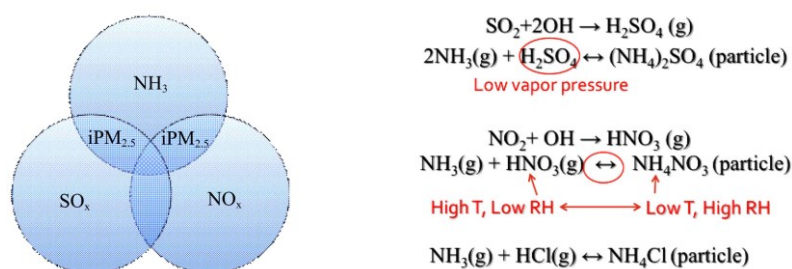


Figure 4 Thermodynamic equilibrium processes of the formation of the secondary iPM<sub>2.5</sub><sup>[11,55]</sup>

Due to the low vapor pressure of H<sub>2</sub>SO<sub>4</sub>, atmospheric NH<sub>3</sub> usually first reacts with H<sub>2</sub>SO<sub>4</sub> to form (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>. If more NH<sub>3</sub> is available, it then reacts with HNO<sub>3</sub> to form (NH<sub>4</sub>NO<sub>3</sub>)<sup>[11]</sup>. These chemical reactions are the most important equilibrium reactions for gas-particle phase partitioning that highly depend on NH<sub>3</sub> and acid concentrations, RH, and temperature<sup>[55,56]</sup>. The reverse reactions occur, especially for HNO<sub>3</sub> under high temperature and low RH such that the equilibrium is shifted to gas phase NH<sub>3</sub> and HNO<sub>3</sub>; vice versa, when the temperature is low and RH is high, the equilibrium is shifted to particle phase of NH<sub>4</sub>NO<sub>3</sub>. Consequently, higher NH<sub>4</sub>NO<sub>3</sub> fraction in winter aerosols is expected<sup>[13,57,58]</sup>. Contributions of inorganic constituents to total PM<sub>2.5</sub> mass demonstrated strong seasonally and geographically variations in response to changes of atmospheric meteorological and chemical conditions.

In addition to the formation of (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> and NH<sub>4</sub>NO<sub>3</sub>, NH<sub>3</sub> may also react with HCl to form NH<sub>4</sub>Cl particle. Contribution of Cl<sup>-</sup> to total inorganic PM<sub>2.5</sub> is usually minor<sup>[13]</sup>.

### 3.2 Ammonia emission inventory and AFO Ammonia emissions

Due to its important impacts on the formation of secondary iPM<sub>2.5</sub> and on the ecosystems in general, AFO

NH<sub>3</sub> emissions have been extensively studied<sup>[59-72]</sup>. Although considerable efforts have been made in developing NH<sub>3</sub> emission inventory that links atmospheric NH<sub>3</sub> to various emission sources<sup>[27,73]</sup>, existing NH<sub>3</sub> emission inventories are considered highly uncertain in assessing significances of various source types. In the U.S., EPA's National Emission Inventory (NEI)<sup>[74]</sup> and CMU (Carnegie Mellon University) NH<sub>3</sub> emission inventory<sup>[75]</sup> are two major NH<sub>3</sub> emission inventory models that have been widely used in various air quality modeling and formation of secondary iPM<sub>2.5</sub> studies. According to EPA's NEI, emissions from AFOs account for approximate 80% of nation's NH<sub>3</sub> emissions (Figure 1). Significance of AFO NH<sub>3</sub> emissions cannot be overlooked in study of formation of iPM<sub>2.5</sub> on local and/or regional scales.

Ammonia has a short lifetime in the atmosphere<sup>[55,76,77]</sup>. Once emitted, NH<sub>3</sub> may be removed from atmosphere by dry and or wet depositions<sup>[78-81]</sup>. Studies of transport of NH<sub>3</sub> and its reaction product NH<sub>4</sub><sup>+</sup> particulate suggest that NH<sub>3</sub> has a short range dispersion due to its fast dry deposition, on the other hand, particulate NH<sub>4</sub><sup>+</sup> may transport in a long a range<sup>[56,79-88]</sup>. Asman and Sutton<sup>[56]</sup> reported that 10% of NH<sub>3</sub> emitted from 1 m height point source was dry deposited within

100 m from the source and 20% within 1 000 m from the source; at a given downwind distance, among the total deposition ( $\text{NH}_3$  plus particulate  $\text{NH}_4^+$ ): 44%  $\text{NH}_3$  dry deposition, 6%  $\text{NH}_3$  wet deposition, 14%  $\text{NH}_4^+$  dry deposition and 36%  $\text{NH}_4^+$  wet deposition. Ammonia dry deposition showed a high spatial variability due to spatial variations in emission sources, land uses and management practices<sup>[56,82-88]</sup>. Fast removal of  $\text{NH}_3$  through dry and wet depositions may limit contribution of agricultural  $\text{NH}_3$  emissions to the formation of secondary  $\text{iPM}_{2.5}$  in the areas far away from agricultural intensive areas where significant  $\text{SO}_2$  or  $\text{NO}_x$  may present in the air (e.g. urban environment).

### 3.3 Thermodynamic equilibrium modeling

As shown in Figure 4, a thermodynamic equilibrium exists between precursor gases and secondary  $\text{NH}_4^+$  particulate of  $\text{iPM}_{2.5}$ . When  $\text{NH}_3$  reacts with acidic gases, it is converted from gaseous phase to particulate phase that is called gas-particle partitioning. Two steps may involve in the gas-particle phase partitioning process. In the first step,  $\text{NO}_x$  and  $\text{SO}_2$  can be transformed into  $\text{HNO}_3$  and  $\text{H}_2\text{SO}_4$  through oxidation processes.  $\text{HNO}_3$  can be produced by heterogeneous hydrolysis of  $\text{N}_2\text{O}_5$ ; hydroxyl radical ( $\text{OH}$ ) can also oxidize  $\text{NO}_x$  into  $\text{HNO}_3$ <sup>[89]</sup>.  $\text{SO}_2$  may be oxidized by oxidants like ozone and hydroxyl radical ( $\text{OH}$ ) into  $\text{H}_2\text{SO}_4$  gas or  $\text{H}_2\text{SO}_4$  ions. In the second step, the three components, i.e.,  $\text{HNO}_3$ ,  $\text{H}_2\text{SO}_4$ ,  $\text{NH}_3$  may partition between gas and particle phases depending on temperature and relative humidity as well as molar concentrations of total sulfate, total nitrate and total ammonia. Nearly all of the  $\text{H}_2\text{SO}_4$  gas can be neutralized and partition to the particle phase due to its low vapor pressure<sup>[90]</sup>.

In gas-particle partitioning processes,  $\text{NH}_3$  preferentially reacts with  $\text{H}_2\text{SO}_4$  to form  $(\text{NH}_4)_2\text{SO}_4$  and  $\text{NH}_4\text{HSO}_4$ , then remaining  $\text{NH}_3$  can react with  $\text{HNO}_3$  to form  $\text{NH}_4\text{NO}_3$ <sup>[89,91]</sup>. Sulfate salts and nitrate salts have different levels of thermal stability characteristics. While sulfate salts are thermally stable<sup>[92]</sup>,  $\text{NH}_4\text{NO}_3$  is not thermally stable enough such that it may decompose to  $\text{HNO}_3$  and  $\text{NH}_3$  when environmental conditions are under high temperature and low relative humidity that do not favor the particle phase partitioning. Potentials for the

formation of sulfate salts and nitrate salts aerosols can be characterized using gas ratio (GR)<sup>[93]</sup>:

$$\text{GR} = \frac{[\text{TA}] - 2[\text{TS}]}{[\text{TN}]}$$

where, [TA] is total amount of  $\text{NH}_3$  and  $\text{NH}_4^+$ ; [TS] is the total amount of  $\text{SO}_4^{2-}$ ,  $\text{HSO}_4^-$  and  $\text{H}_2\text{SO}_4$ ; [TN] is the total amount of  $\text{NO}_3^-$  and  $\text{HNO}_3$ .

In  $\text{NH}_3$  poor regime,  $\text{GR} < 0$ , this is insufficient  $\text{NH}_3$  to neutralize all  $\text{SO}_4^{2-}$ ; in moderate regime,  $0 < \text{GR} < 1$ , there is sufficient  $\text{NH}_3$  to neutralize  $\text{SO}_4^{2-}$  but not  $\text{NO}_3^-$  in  $\text{NH}$ -rich regime,  $\text{GR} > 1$ , there is sufficient  $\text{NH}_3$  to neutralize both  $\text{SO}_4^{2-}$  and  $\text{NO}_3^-$ . According to Wu et al.<sup>[94]</sup> in winter condition,  $\text{NH}_3$  may not be fully neutralized by  $\text{SO}_4^{2-}$ , a more generic term  $\text{NH}_3 + \text{NO}_3^-$  should be used to account for neutralization by  $\text{NO}_3^-$  such that an adjusted GR (AdjGR) should be used to examine  $\text{iPM}_{2.5}$  sensitivity to  $\text{NH}_3$  emission/ concentration.

$$\text{AdjGR} = \frac{[\text{NH}_3] + [\text{NO}_3^-]}{[\text{TN}]}$$

In addition to GR examination, the molar ratios of  $(\text{NH}_4^+)/\text{SO}_4^{2-}$  (MR) were used to estimate neutralization of  $\text{NH}_3$  associated with  $\text{SO}_4^{2-}$ <sup>[13,36,37,95]</sup>.

$$\text{MR} = \frac{[\text{NH}_4^+] - [\text{NO}_3^-] - [\text{Cl}^-]}{[\text{SO}_4^{2-}]}$$

where,  $[\text{NH}_4^+]$ ,  $[\text{NO}_3^-]$ ,  $[\text{Cl}^-]$  and  $[\text{SO}_4^{2-}]$  are measured molar concentrations of  $\text{NH}_4^+$ ,  $\text{NO}_3^-$ ,  $\text{Cl}^-$  and  $\text{SO}_4^{2-}$ , respectively. Formation of various  $\text{iPM}_{2.5}$  constituents would follow the pattern below<sup>[93]</sup>:

- 1) When  $\text{MR} < 0.5$ , aerosol phase consist of  $\text{H}_2\text{SO}_4$  and  $\text{NH}_4\text{HSO}_4$ ;
- 2) When  $0.5 < \text{MR} < 1.25$ , aerosol phase is dominant by  $\text{NH}_4\text{HSO}_4$ ;
- 3) When  $\text{MR} = 1.25$ , aerosol phase is dominant by  $(\text{NH}_4)_3\text{H}(\text{SO}_4)_2$ ;
- 4) When  $\text{MR} = 1.5$ , aerosol phase consists of  $(\text{NH}_4)_3\text{H}(\text{SO}_4)_2$ ;
- 5) When  $\text{MR} > 1.5$ , formation of  $(\text{NH}_4)_2\text{SO}_4$ ;
- 6) When  $\text{MR} = 2$ , complete neutralization of sulfate and predominance of  $(\text{NH}_4)_2\text{SO}_4$ .

Given the elevated concentration of  $\text{NH}_3$  around AFO production facilities, the MR in AFO environments is expected to be 2. However, the observation by Li et al.<sup>[36,37]</sup> indicated that wind speed may affect transport of

NH<sub>3</sub> within the time scale of establishment of local gas-particle phase equilibrium such that low (NH<sub>4</sub><sup>+</sup>)<sub>s</sub>/SO<sub>4</sub><sup>2-</sup> molar ratios (<2) may occur indicating gas phase NH<sub>3</sub> was not in equilibrium with sulfate to form (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>. Impact of wind speed, direction needs to be further investigated.

In study of gas-particle phase partitioning, a thermodynamic equilibrium model ISORROPIA-II was commonly used to examine response of iPM<sub>2.5</sub> to the change of precursor gas as well as to the changes of meteorological conditions<sup>[13,36,37,89,96]</sup>. The model inputs included simultaneously measured concentrations of NH<sub>3</sub>, NO<sub>x</sub>, SO<sub>2</sub>, SO<sub>4</sub><sup>2-</sup>, NO<sub>3</sub><sup>-</sup>, Cl<sup>-</sup>, Ca<sup>+</sup>, K<sup>+</sup>, RH, T, and the model outputs included total aerosol NH<sub>4</sub><sup>+</sup>, total aerosol NO<sub>3</sub><sup>-</sup>, total aerosol SO<sub>4</sub><sup>2-</sup>, total aerosol Cl<sup>-</sup>, aerosol aqueous phase mass, aerosol solid phase mass. Simulation of iPM<sub>2.5</sub> responses to reductions in precursor concentrations and T/RH was also conducted to address dynamics of the formation of iPM<sub>2.5</sub><sup>[36,37]</sup>.

#### 4 Contributions of AFO NH<sub>3</sub> emissions to the secondary iPM<sub>2.5</sub>

Although the fundamental knowledge about the formation of secondary NH<sub>4</sub><sup>+</sup> particulate is well established, the contributions of AFO air emissions to atmospheric secondary iPM<sub>2.5</sub> have not been well studied and experimentally quantified. Based upon the amount of NH<sub>4</sub><sup>+</sup> in sulfate and nitrate PM<sub>2.5</sub> from CASTNET and an assumption that the amount of NH<sub>4</sub><sup>+</sup> in sulfate and nitrate PM<sub>2.5</sub> emitted from AFOs is in the same fraction as that of the total NH<sub>3</sub> emissions attributable to AFOs, Hristov<sup>[97]</sup> conducted a hypothetical study to assess contribution of AFO NH<sub>3</sub> emissions to secondary iPM<sub>2.5</sub>. It was discovered that iPM<sub>2.5</sub> formed from AFO NH<sub>3</sub> emissions contributed 5%-11% of total PM<sub>2.5</sub> mass across different regions in the U.S. and under different weather conditions. Under cold weather conditions, this contribution may reach as high as 20% in eastern U.S. For several reasons, research findings from this study are challenges and remain to be tested. First of all, chemical speciation data from CASTNET are not applicable to the AFO environment since the PM<sub>2.5</sub> samples were not taken from AFO environment. Secondly, there was lack of

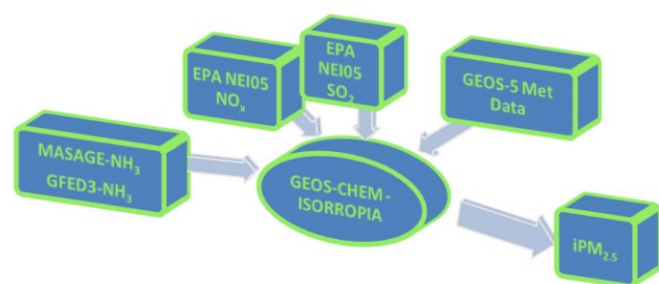
consideration of spatial and temporal variations in NH<sub>3</sub> emission inventory used in the study. Thirdly, there was lack of consideration of acidic gas availability in assessing the formation of secondary iPM<sub>2.5</sub>. Last, but not least, there was lack of model validation in application of the hypothetical models to quantify the formation of iPM<sub>2.5</sub>.

Walker et al.<sup>[13,96]</sup> conducted several field monitoring studies coupled with statistic and ISORROPIA modeling to assess response atmospheric iPM<sub>2.5</sub> to precursor gas emissions at agricultural sites in southeastern U.S. where animal production and fertilizer are major sources of NH<sub>3</sub> emissions. It was revealed that NH<sub>4</sub><sup>+</sup>, SO<sub>4</sub><sup>2-</sup>, NO<sub>3</sub><sup>-</sup> and Cl<sup>-</sup> accounted for 22%, 53%, 24%, and 1% of total iPM<sub>2.5</sub> mass, respectively. Reductions of SO<sub>2</sub> and NO<sub>x</sub> will lead to reduction of SO<sub>4</sub><sup>2-</sup> and NO<sub>3</sub><sup>-</sup> associated iPM<sub>2.5</sub> as well as total PM<sub>2.5</sub>. Reducing SO<sub>2</sub> emissions is more effective in summer; whereas reducing NO<sub>x</sub> and/or NH<sub>3</sub> emissions is more effective in winter time, which may lead to the greatest reduction of iPM<sub>2.5</sub>. In agricultural areas where high NH<sub>3</sub> emissions are produced, NH<sub>4</sub><sup>+</sup> particulate formation is less likely to be limited by NH<sub>3</sub> availability, but more likely to be limited by availability of SO<sub>2</sub> and NO<sub>x</sub>. Response of iPM<sub>2.5</sub> to NH<sub>3</sub> concentrations/emissions is affected by the concentrations of acidic gas species, i.e. SO<sub>2</sub> and NO<sub>x</sub>. When the acidic gas species are limited, significant reduction of NH<sub>3</sub> emissions/concentrations may not be effective in reducing iPM<sub>2.5</sub> concentrations.

In a recent field study by Li et al.<sup>[32,36,37,51]</sup>, NH<sub>3</sub> concentrations and inorganic species of PM<sub>2.5</sub> in the vicinity of an AFO were simultaneously measured to investigate formation of secondary iPM<sub>2.5</sub>. Response of NH<sub>4</sub><sup>+</sup> particulate to precursor gas concentrations and environmental parameters was modeled by ISORROPIA-II, using measured gas and particle concentrations as well as environmental parameters. It was discovered that PM<sub>2.5</sub> level responded nonlinearly to changes in NH<sub>3</sub> concentrations. While there was abundant NH<sub>3</sub> in the vicinity of this AFO facility, PM<sub>2.5</sub> concentration was relatively insensitive to changes in NH<sub>3</sub>. The ISORROPIA results indicated that adding more NH<sub>3</sub> caused gas phase NH<sub>3</sub> to increase linearly but

$\text{NH}_4^+$  particulate leveled off as total  $\text{NH}_3$  reached to a level greater than  $10 \mu\text{g}/\text{m}^3$ . The model results also showed that  $\text{NH}_3$  first reacted with  $\text{H}_2\text{SO}_4$ , then the remaining  $\text{NH}_3$  would react with  $\text{HCl}$  and  $\text{HNO}_3$ , depending on the RH and temperature, therefore, the reduction of  $\text{SO}_4^{2-}$  had the most significant impact on the reduction of  $\text{PM}_{2.5}$ , and the abundant  $\text{NH}_3$  in the system will react with  $\text{H}_2\text{SO}_4$  to form  $(\text{NH}_3)_2\text{SO}_4$ . A reduction in  $\text{NH}_3$  had limited impact on inorganic  $\text{PM}_{2.5}$  at the research site due to limited concentrations of acidic gases.

Taking modeling approach, Paulot and Jacob<sup>[7]</sup> investigate the impact of  $\text{NH}_3$  emissions from US food export on atmospheric  $\text{PM}_{2.5}$ . Figure 5 illustrates the flow diagram of this modeling practice. It was reported that on average, the US food export increases  $\text{PM}_{2.5}$  exposure by  $0.36 \mu\text{g}/\text{m}^3$ . Due to lack of model validation, findings from this research remain to be tested and verified with observation data.



Note: MASAGE: model for agricultural  $\text{NH}_3$  emission; GFED3: model for other anthropogenic sources  $\text{NH}_3$  emissions; NEI05: national emission inventory for 2005; GEOS-5: NASA Goddard Earth Observation System for meteorological data

Figure 5 Modeling flow diagram for assessing the impact of  $\text{NH}_3$  emissions from food export on  $i\text{PM}_{2.5}$  by Paulot and Jacob<sup>[7]</sup>

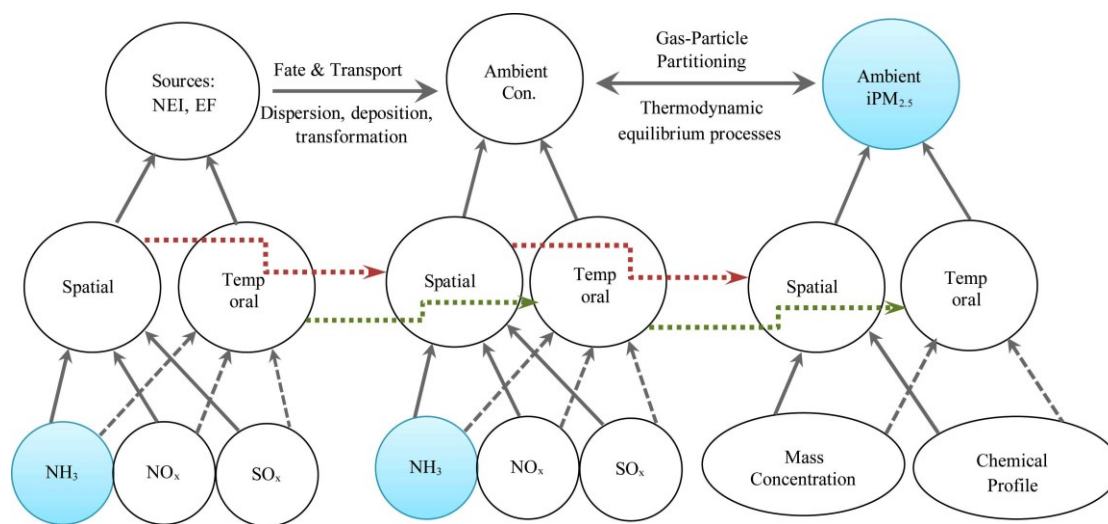
While only limited research has been done on quantifying responses of the atmospheric secondary  $i\text{PM}_{2.5}$  to AFO  $\text{NH}_3$  emissions<sup>[13,32,36,37,51,52,79,94,96]</sup>, impact of fate and transport of AFO  $\text{NH}_3$  emissions on the formation of ambient  $i\text{PM}_{2.5}$  is not truly understood. There is an urgent need for advanced understanding of fate and transport of AFO  $\text{NH}_3$  emissions and how it would affect its potential for contributing to the formation of secondary  $\text{PM}_{2.5}$  through experimental evaluations in the field and/or through simulation and validation of current atmospheric transport models and/or thermodynamic equilibrium aerosol model under the AFO environments<sup>[98]</sup>.

## 5 Summary and recommendations

Animal feeding operations have been perceived to be important contributors to atmospheric  $\text{PM}_{2.5}$ . Although knowledge about PM chemical compositions is essential for source identification and development of source-specific PM mitigation techniques, chemical source profiles of AFO PM and chemical compositions of PM inside AFO facilities and in the surrounding areas of the facilities are unknown. Several ambient chemical speciation networks (e.g. IMPROVE, CASTNET, CSN) provide high quality PM chemical speciation datasets for air quality and public health studies, however, there is a lack of representation of intensive agricultural operation areas in networks to assess ambient  $i\text{PM}_{2.5}$  as impacted by AFO  $\text{NH}_3$  emissions that accounts for more than half of nation's  $\text{NH}_3$  emissions.

Great challenge exists in assessing contributions of AFO  $\text{NH}_3$  emissions to secondary  $i\text{PM}_{2.5}$  due to lack of temporal and spatial monitoring data of atmospheric  $\text{SO}_2$ ,  $\text{NO}_x$  and  $\text{NH}_3$  concentrations in agricultural production areas where most  $\text{NH}_3$  emissions are generated. Moreover, lack of holistic understanding of the pathway from  $\text{NH}_3$  emissions to atmospheric  $i\text{PM}_{2.5}$  (Figure 6) also limits our understanding of impact of  $\text{NH}_3$  emissions on the formation of secondary  $i\text{PM}_{2.5}$  at regional scale. Research gaps remain in systematic understanding of (1)  $\text{NH}_3$  spatial and temporal variations of precursor gas emissions (i.e.  $\text{NH}_3$ ,  $\text{NO}_x$ ,  $\text{SO}_x$ ); (2) deposition and transport of  $\text{NH}_3$ ,  $\text{NO}_x$ ,  $\text{SO}_x$  as impacted by emission source characteristics, land uses and management practices; (3) Dynamics of gas-particle phases ( $\text{NH}_3/\text{NH}_4^+$ ) partitioning in response to the spatial and temporal variations of atmospheric  $\text{NH}_3$ ,  $\text{NO}_x$ ,  $\text{SO}_x$  concentrations as well as meteorological conditions. It is likely that fast removal of  $\text{NH}_3$  through dry and wet deposition may limit agricultural  $\text{NH}_3$  emission contribution to the formation of secondary  $i\text{PM}_{2.5}$  in the area far away from emission sources where significant  $\text{SO}_2$  or  $\text{NO}_x$  may present in the air. Further investigation is needed to advance our understanding of ambient secondary  $i\text{PM}_{2.5}$  as impacted by agricultural  $\text{NH}_3$  emissions and various ammonia mitigation strategies.





Note: NEI: National Emission Inventory, EF: Emission Factor.

Figure 6 Holistic pathways to the formation of atmospheric iPM<sub>2.5</sub>

Based upon this comprehensive literature review of previous research on iPM<sub>2.5</sub> formation as impacted by AFO air emissions, following research needs are identified:

- Continued improvement of AFO NH<sub>3</sub> emission inventory;
- Simultaneous measurements of precursor gases (e.g. NH<sub>3</sub>, NO<sub>x</sub>, SO<sub>2</sub>, etc.), PM<sub>2.5</sub> chemical compositions, and meteorological conditions in intensive AFO production areas to assess formation of secondary iPM<sub>2.5</sub> as impacted by AFO NH<sub>3</sub> emissions, and atmospheric chemical and meteorological conditions;
- Experimental based assessment of NH<sub>3</sub> and ammonium (NH<sub>4</sub><sup>+</sup>) particulate dry deposition rates and velocities as impacted by AFO NH<sub>3</sub> emissions, distance from emission source, land usage, and meteorological conditions;
- Experimental identification of chemical compositions of AFO PM, both inside the AFO production facilities and in the vicinity of the those facilities;
- Development of experimental-based models describing NH<sub>3</sub>/NH<sub>4</sub> deposition and formation of iPM<sub>2.5</sub> as a function of house emission rates, distance from the sources, ambient meteorological (e.g. T, RH, wind speed/direction), and chemical (e.g. SO<sub>2</sub>, NO<sub>x</sub>, etc.) conditions;
- Experimental validation of atmospheric transport models (e.g. CMAQ, etc.) for modelling NH<sub>3</sub> deposition in intensive AFO areas;

- Experimental validation of atmospheric transport models (e.g. CMAQ, etc.) and thermodynamic equilibrium aerosol models (e.g., ISORROPIA) for modelling gas-particle phase partitioning of secondary iPM<sub>2.5</sub> in AFO environments;
- Experimental based identification of key factors that affect precursor gas concentrations, and formation of secondary iPM<sub>2.5</sub> in AFO environments;
- Assessment of impact of management practices and mitigation strategies on NH<sub>3</sub> emissions, fate and transport of the emissions, and formation of secondary iPM<sub>2.5</sub>.

## [References]

- [1] US EPA. Regulatory Impact Analysis for the Final Revisions to the National Ambient Air Quality Standards for Particulate Matter. 2012. <http://www.epa.gov/ttn/ecas/regdata/RIAs/finalria.pdf>. Accessed on [2015-03-12].
- [2] USEPA. National Emission Inventory-National Emissions Inventory, version 1, technical support document. Revised draft report. U. S. Environmental Protection Agency, Washington, DC. 2013.
- [3] Donham K J, Reynolds S J, Whitten P, Merchant J A, Burmeister L, Pependorf W J. Respiratory dysfunction in swine production facility workers: dose-response relationships of environmental exposures and pulmonary-function. *American Journal of Industrial Medicine*, 1995; 27(3): 405-418.
- [4] Pope III C A, Ezzati M, Dockery D W. Fine-particulate air pollution and life expectancy in the United States. *New England Journal of Medicine*, 2009; 360(4): 376-386.
- [5] Pui D Y H, Chen S C, Zuo Z L. PM<sub>2.5</sub> in China:

- Measurements, sources, visibility and health effects, and mitigation. *Particology*, 2014; 13: 1–26.
- [6] Franklin M, Koutrakis P, Schwartz J. The role of particle composition on the association between PM<sub>2.5</sub> and mortality. *Epidemiology (Cambridge, Mass.)*, 2008; 19(5): 680–689.
- [7] Paulot F, Jacob D J. Hidden cost of U.S. agricultural exports: particulate matter from ammonia emissions. *Environmental Science and Technology*, 2014; 48(2), 903–908.
- [8] US EPA. Visibility in Mandatory Federal Class I Areas, 1994-1998. 1998. <http://www.epa.gov/airquality/visibility/report/index.html>. Accessed on [2015-03-12]
- [9] US EPA. Integrated science assessment for particulate matter (final report). U.S. Environmental Protection Agency Washington, DC, 2009.
- [10] Hinds WC. *Aerosol Technology; Properties, Behavior and Measurement of Airborne Particles*. New York: John Wiley and Sons, 1998.
- [11] Seinfeld J H, Pandis S N. *Atmospheric Chemistry and Physics: From Air Pollution to Climate Change*. New York: John Wiley and Sons, 1998.
- [12] NARSTO. *Particulate Matter Science for Policymakers: A NARSTO Assessment*. UK: Cambridge University Press, 2004.
- [13] Walker J T, Robarge W P, Shendrikar A, Kimball H. Inorganic PM<sub>2.5</sub> at a U.S. agricultural site. *Environmental Pollution*, 2006; 139(2): 258–271.
- [14] Malm W C, Sisler J F, Huffman D, Eldred R A, Cahill T S. Spatial and seasonal trends in particle concentration and optical extinction in the United States. *Journal of Geophysical Research: Atmospheres (1984–2012)*, 1994; 99(D1): 1347–1370.
- [15] Tolocka M P, Solomon P A, Mitchell W, Norris G A, Gemmill D B, Weiner R W, et al. East versus West in the US: chemical characteristics of PM<sub>2.5</sub> during the winter of 1999. *Aerosol Science and Technology*, 2001; 34(1): 88–96.
- [16] Henze D K, Seinfeld J H, Shindell D T. Inverse modeling and mapping US air quality influences of inorganic PM<sub>2.5</sub> precursor emissions using the adjoint of GEOS-Shem. *Atmospheric Chemistry and Physics*, 2009; 9(16): 5877–5903.
- [17] Heald C L, Collett Jr J L, Lee T, Benedict K B, Schwandner F M, Li Y, et al. Atmospheric ammonia and particulate inorganic nitrogen over the United States. *Atmospheric Chemistry and Physics*, 2012; 12(21): 10295–10312.
- [18] Ianniello A, Spataro F, Esposito G, Allegrini I, Hu M, Zhu T. Chemical characteristics of inorganic ammonium salts in PM<sub>2.5</sub> in the atmosphere in Beijing (China). *Atmospheric Chemistry and Physics*, 2011; 11(21): 10803–10822, 2011.
- [19] Wang Y, Zhuang G, Tang A, Yuan H, Sun Y, Chen S, Zheng A. The ion chemistry and the source of PM<sub>2.5</sub> aerosol in Beijing. *Atmospheric Environment*, 2005; 39(21): 3771–3784.
- [20] Sun Y L, Zhuang G S, Wang Y, Han L H, Guo J H, Dan M, et al. The air-borne particulate pollution in Beijing – concentration, composition, distribution and sources. *Atmospheric Environment*, 2004; 38(35): 5991–6004.
- [21] Yao X, Chan C K, Fang M, Cadle S, Chan T, Mulawa P, et al. The water-soluble ionic composition of PM<sub>2.5</sub> in Shanghai and Beijing, China. *Atmospheric Environment*, 2002, 36(26): 4223–4234.
- [22] Pinder R W, Adams P J, Pandis S N. Ammonia emission controls as a cost-effective strategy for reducing atmospheric particulate matter in the Eastern United States. *Environmental Science and Technology*, 2007, 41(2): 380–386.
- [23] US EPA. Estimating ammonia emissions from anthropogenic nonagricultural sources - draft final report. 2004. [http://www.epa.gov/ttn/chief/eiip/techreport/volume03/eiip\\_areasourcesnh3.pdf](http://www.epa.gov/ttn/chief/eiip/techreport/volume03/eiip_areasourcesnh3.pdf). Accessed on [2015-03-14].
- [24] US EPA. Air emission characterization and management. Supporting documentation for the EPA regional science workshop on animal feeding operations (AFOs) – science and technical support needs, 2004. [http://www.epa.gov/osp/presentations/afo/s2\\_airemis.pdf](http://www.epa.gov/osp/presentations/afo/s2_airemis.pdf). Accessed on [2015-03-14].
- [25] US EPA. Air Quality Compliance Agreement for Animal Feeding Operations. 2005. <http://www.epa.gov/compliance/resources/agreements/caa/caf0-agr-0501.html>. Accessed on [2015-03-14].
- [26] US EPA. Agriculture-Air Monitoring, Data from Sites Monitored. 2010. <http://www.epa.gov/airquality/agmonitoring/data.html>. Accessed on [2014-07-07].
- [27] NRC. *Air Emissions from Animal Feeding Operations: Current Knowledge, Future Needs*. US: Washington D.C, The National Academies Press, 2003.
- [28] Cambra-López M, Aarnink A J A, Zhao Y, Calvet Y, Torres A G. Airborne particulate matter from livestock production systems: a review of an air pollution problem. *Environmental Pollution*, 2010; 158(1): 1–17.
- [29] Cambra-López M, Torres A G, Aarnink A J A, Ogink N W M. Source analysis of fine and coarse particulate matter from livestock houses. *Atmospheric Environment*, 2011; 45(3): 694–707.
- [30] Cambra-López M, Hermosilla T, Lai H T L, Aarnink A J A, Ogink N W M. Particulate matter emitted from poultry and pig houses: source identification and quantification. *Transaction of the ASABE*, 2011; 54(2): 629–642.
- [31] Yang X, Wang X, Zhang Y, Lee J, Su J, Gates R S. Characterization of trace elements and ions in PM<sub>10</sub> and

- PM<sub>2.5</sub> emitted from animal confinement buildings. *Atmospheric Environment*, 2011; 45(39): 7096–7104.
- [32] Li Q F. Particulate matter from an egg production facility: emission, chemistry and local dispersion. PhD dissertation. Raleigh: North Carolina State University, 2012.
- [33] Wang-Li L, Cao Z, Buser M, Whitelock D, Parnell C B, Zhang Y. Techniques for measuring particle size distribution of particulate matter emitted from animal feeding operations. *Journal of Atmospheric Environment*, 2013; 66: 25–32.
- [34] Wang-Li L, Cao Z, Li Q, Liu Z, Beasley D B. Concentration and particle size distribution of particulate matter emitted from tunnel ventilated high-rise layer operation houses. *Atmospheric Environment*, 2013, 66: 8–16.
- [35] Finlayson-Pitts B J, Pitts J N. *Atmospheric Chemistry: Fundamentals and experimental techniques*. New York: John Wiley and Sons, 1986.
- [36] Li Q F, Wang-Li L, Liu Z, Jayanty R K M., Shah S B, Bloomfield P. Major ionic compositions of fine particulate matter in an animal feeding operation facility and its vicinity. *Journal of the Air and Waste Management Association*, 2014; 64(11): 1279–1287.
- [37] Li Q F, Wang-Li L, Shah S B, Jayanty R K M, Bloomfield P. Ammonia concentrations and modeling inorganic particulate matter in the vicinity of an egg production facility. *Journal of Environmental Science and Pollutant Research*, 2014; 21(6): 4675–4685.
- [38] Li Q F, Wang-Li L, Jayanty R K M, Shah S B. Elemental Composition and Chemical Mass Closure of Fine Particulate in an Animal Feeding Operation Facility and its Vicinity. *Journal of Environmental Protection*. Accepted.
- [39] Liang J, Jacobson M Z. A study of sulfur dioxide oxidation pathways over a range of liquid water contents, pH values, and temperatures. *Journal of Geophysical Research: Atmospheres*, 1999; 104(D11): 13749–13769.
- [40] US EPA. Chemical Speciation-Laboratory Standard Operation Procedures (SOPs), 2011. <http://www.epa.gov/ttnamti1/specsop.html>. Accessed on [2015-03-12].
- [41] Wang-Li L. Techniques for characterization of particulate matter emitted from animal feeding operations. Peer-reviewed book chapter of “Evaluating Veterinary Pharmaceutical Behavior in the Environments”. American Chemical Society (ACS). 2013; pp15–39. DOI: 10.1021/bk-2013-1126.ch002.
- [42] IMPROVE. Interagency Monitoring of Protected Visual Environments. <http://vista.cira.colostate.edu/improve/>. Accessed on [2015-03-12].
- [43] US EPA. [www.epa.gov/CASTNET](http://www.epa.gov/CASTNET). Accessed on [2015-03-12].
- [44] US EPA. Chemical Speciation. <http://www.epa.gov/ttn/amtic/speciepg.html>. Accessed on [2015-03-12].
- [45] Jayanty R K M, Flanagan J B. An overview of chemical speciation fine particle monitoring networks in the U.S.A. 2010. [http://www.dri.edu/images/stories/editors/leapfrog/techprog/Ie\\_6\\_Jayanty.pdf](http://www.dri.edu/images/stories/editors/leapfrog/techprog/Ie_6_Jayanty.pdf). Accessed on [2015-03-14].
- [46] US EPA, CASTNET 2012 Annual Report. 2012. [http://epa.gov/castnet/javaweb/docs/annual\\_report\\_2012.pdf](http://epa.gov/castnet/javaweb/docs/annual_report_2012.pdf). Accessed on [2015-03-12].
- [47] Aarnink A J A, Roelofs P F M M, Ellen H H, Gunnink H. Dust sources in animal houses. 1999.
- [48] Cambra-López M, Hermosilla T, Aarnink A J A, Ognik N. Selection of particle characteristics to distinguish amongst potential sources of particulate matter in poultry and pigs. An ASABE Meeting Presentation, 2011. doi: 10.13031/2013.37777
- [49] Yang X, Wang X; Zhang Y, Lee J, Su J; Gates R S. Characterization of trace elements and ions in PM<sub>10</sub> and PM<sub>2.5</sub> emitted from animal confinement buildings. *Atmospheric Environment*, 2011; 45(39): 7096–7104.
- [50] Zhao Y, Cliff S S, Wexler A S, Javed W, Perry K, Pan Y, et al. Measurements of size- and time-resolved elemental concentration at a California dairy farm. *Atmospheric Environment*, 2014, 94: 773–781.
- [51] Li Q F, Wang-Li L, Liu Z F, Beasley D B. Chemical characterization of particulate matter emitted from animal feeding operations. ASABE Technical Paper, 2009.
- [52] Li QF, Wang-Li L, Jayanty R K M, Shah S B. Organic and elemental carbons in atmospheric fine particulate in an animal agriculture intensive area in North Carolina: estimation of secondary organic carbon concentration. *Open Journal of Air Pollution*, 2013; 2: 7–18.
- [53] Saxena P, Hudischewskyj A B, Seigneur C, Seinfeld J H. A comparative study of equilibrium approaches to the chemical characterization of secondary aerosols. *Atmospheric Environment*, 1986; 20(7): 1471–1483.
- [54] Tanner R L, Marlow W H, Newman L. Chemical composition correlations of size-fractionated sulfate in New York City aerosol. *Environmental Science and Technology*, 1979; 13(1): 75–78.
- [55] Zhang Y, Wu S Y, Krishnan S, Wang K, Queen A, Aneja V P, et al. Modeling agricultural air quality: current status, major challenges and outlook. *Atmospheric Environment*, 2008; 42(14): 3218–3237.
- [56] Asman W A H, Sutton M A, Schjørring J K. Ammonia: emission, atmospheric transport and deposition. *New Phytologist*, 1998; 139(1): 27–48.
- [57] Stelson A W, Seinfeld J H. Relative humidity and temperature dependence of the ammonium nitrate dissociation constant. *Atmospheric Environment*, 1982;

- 16(5): 983–992.
- [58] Mozurkewich M. The dissociation constant of ammonium nitrate and its dependence on temperature, relative humidity and particle size. *Atmospheric Environment. Part A. General Topics*, 1993; 27(2): 261–270.
- [59] Wang-Li L, Li Q F, Wang K, Bogan B W, Ni J Q, Cortus E L, et al. National air emissions monitoring study's Southeast layer site: Part I. site characteristics and monitoring methodology. *Transaction of the ASABE*, 2013; 56(3): 1157–1171.
- [60] Li Q F, Wang-Li L, Wang K, Chai L, Cortus E L, Kilic I, et al. National air emissions monitoring study's Southeast layer site: Part II. particulate matter. *Transaction of the ASABE*. 2013; 56(3): 1173–1184.
- [61] Wang-Li L, Li Q F, Chai L, Cortus E L, Wang K, Kilic I, et al. National air emissions monitoring study's Southeast layer site: Part III. ammonia concentrations and emissions. *Transaction of the ASABE*, 2013; 56(3): 1185–1197.
- [62] Li Q F, Wang-Li L, Bogan B W, Wang K, Chai L, Ni J Q, et al. National air emissions monitoring study's Southeast layer site: Part IV. effects of farm management. *Transaction of the ASABE*, 2013; 56(3): 1199–1209.
- [63] Arogo J, Westerman P W, Heber A J, Robarge W P, Classen J J. Ammonia emissions from animal feeding operations. *National Center for Manure and Animal Waste Management White Papers*, 2002; 41–88.
- [64] Faulkner W B, Shaw B W. Review of ammonia emission factors from United States animal agriculture. *Atmospheric Environment*, 2008, 42(27): 6565–6574.
- [65] Gates R S, Casey K D, Wheeler E F, Xin H, Pescatore A J. U.S. broiler housing ammonia emissions inventory. *Atmospheric Environment*, 2008; 42(14): 3342–3350.
- [66] Lin X J, Cortus E L, Zhang R, Jiang S, Heber A J. Ammonia, hydrogen sulfide, carbon dioxide, and particulate matter emissions from California high-rise layer houses. *Atmospheric Environment*, 2012; 46: 81–91.
- [67] Maghirang R G, Manbeck H B, Roush W B, Muir F V. Air contaminant distributions in a commercial laying house. *Transactions of the ASAE (USA)*, 1991; 34(5): 2171–2179.
- [68] Ni J Q, Chai L, Chen L, Bogan B W, Wang K, Cortus E L, et al. Characteristics of ammonia, hydrogen sulfide, carbon dioxide, and particulate matter concentrations in high-rise and manure-belt layer hen houses. *Atmospheric Environment*, 2012; 57: 165–174.
- [69] Nicholson F A, Chambers B J, Walker A W. Ammonia emissions from broiler litter and laying hen manure management systems. *Biosystems Engineering*, 2004; 89(2): 175–185.
- [70] Roumeliotis T S, Dixon B J, Van Heyst B J. Characterization of gaseous pollutant and particulate matter emission rates from a commercial broiler operation: Part II. Correlated emission rates. *Atmospheric Environment*, 2010; 44(31): 3778–3786.
- [71] Roumeliotis T S, Dixon B J, Van Heyst B J. Characterization of gaseous pollutant and particulate matter emission rates from a commercial broiler operation: Part I. Observed trends in emissions. *Atmospheric Environment*, 2010; 44(31): 3770–3777.
- [72] Wheeler E F, Casey K D, Gates R S, Xin H, Zajackowski J L, Topper P A, et al. Ammonia emissions from twelve U.S. broiler chicken houses. *Transactions of the ASAE*, 2006; 49(5): 1495–1512.
- [73] Pinder R W, Adams P J, Pandis S N, Gilliland A B. Temporally resolved ammonia emission inventories: Current estimates, evaluation tools, and measurement needs. *Journal of Geophysical Research: Atmospheres*, 2006, 111(D16): 310.
- [74] US EPA. The 2011 National emissions inventory. <http://www.epa.gov/ttnchie1/net/2011inventory.html>. Accessed on [2015-03-14].
- [75] Strader R, Peckney N J, Pinder R W, Adams P J, Goebes M, Ayers J, et al. The CMU ammonia emission inventory, 2005. Available at <http://www.cmu.edu/ammonia>. Accessed on March 15, 2015.
- [76] Adams P J, Seinfeld J H, Koch D M. Global concentrations of tropospheric sulfate, nitrate, and ammonium aerosol simulated in a general circulation model. *Journal of Geophysical Research: Atmospheres*, 1999, 104(D11): 13791–13823.
- [77] Aneja V P, Roelle P A, Murray G C, Southerland J, Erisman J W, Fowler D, et al. Atmospheric nitrogen compounds II: emissions, transport, deposition, and assessment. *Atmospheric Environment*, 2001; 35(11): 1903–1911. doi: 10.1016/S1352-2310(00)00543-4.
- [78] NADP. Ammonia monitoring network (AMoN). <http://nadp.sws.uiuc.edu/amon/>. Accessed on [2015-03-14].
- [79] Aneja V P, Nelson D R, Roelle P A, Walker J T. Agricultural ammonia emissions and ammonium concentrations associated with aerosols and precipitation in southeast United States. *Journal of Geophysical Research: Atmospheres (1984–2012)*, 2003; 108(D4): 4152.
- [80] Walker J T, Aneja V P, Dickey D A. Atmospheric transport and wet deposition of ammonium in North Carolina. *Atmospheric Environment*, 2000; 34(20): 3407–3418.
- [81] Walker J, Nelson D, Aneja V P. Trends in ammonium concentration in precipitation and atmospheric ammonia emissions at a coastal plain site in North Carolina U.S.A. *Environmental Science and Technology*, 2000; 34(17): 3527–3534.
- [82] Duyzer J. Dry deposition of ammonia and ammonium

- aerosols over heathland. *Journal of Geophysical Research: Atmospheres*, 1994; 99(D9): 18757–18763.
- [83] Sutton M A, Asman W A H, Schjoerring J K. Dry deposition of reduced nitrogen. *Tellus B*, 1994; 46(4): 255–273.
- [84] Asman W A H. Factors influencing local dry deposition of gases with special reference to ammonia. *Atmospheric Environment*, 1998; 32(3): 415–421.
- [85] Asman W A H. Modelling the atmospheric transport and deposition of ammonia and ammonium: an overview with special reference to Denmark. *Atmospheric Environment*, 2001; 35(11): 1969–1983.
- [86] Asman W A H, Drukker B, Janssen A J. Modelled historical concentrations and depositions of ammonia and ammonium in Europe. *Atmospheric Environment*, 1988; 22(4): 725–735.
- [87] Asman W A H, Harrison R M, Ottley C J. Estimation of the net air-sea flux of ammonia over the southern bight of the North Sea. *Atmospheric Environment*, 1994; 28(22): 3647–3654.
- [88] Asman W A H, Janssen A J. A long-range transport model for ammonia and ammonium for Europe. *Atmospheric Environment*, 1987; 21(10): 2099–2119.
- [89] Nenes A, Pandis S N, Pilinis C. ISORROPIA: a new thermodynamic equilibrium model for multiphase multicomponent inorganic aerosols. *Aquatic Geochemistry*, 1998; 4(1): 123–152.
- [90] Makar P A, Moran M D, Zheng Q, Cousineau S, Sassi M, Duhamel A, et al. Modeling the impacts of ammonia emissions reductions on North American air quality, *Atmospheric Chemistry and Physics Discussions*, 2009; 9(2): 5371–5422.
- [91] Huntzicker J J, Cary R A, Ling C S. Neutralization of sulfuric acid aerosol by ammonia. *Environmental Science and Technology*, 1980; 14(7): 819–824.
- [92] Olszyna K J, Bairai S T, Tanner R L. Effect of ambient NH<sub>3</sub> levels on PM<sub>2.5</sub> composition in the Great Smoky Mountains National Park. *Atmospheric Environment*, 2005, 39(25): 4593–4606.
- [93] Ansari A S, Pandis S N. An analysis of four models predicting the partitioning of semivolatile inorganic aerosol components. *Aerosol Science and Technology*, 1999, 31(2-3): 129–153.
- [94] Wu S Y, Hu J L, Zhang Y, Aneja V P. Modeling atmospheric transport and fate of ammonia in North Carolina—Part II: Effect of ammonia emissions on fine particulate matter formation. *Atmospheric Environment*, 2008, 42(14): 3437–3451.
- [95] Liu Z, Wang L, Li Q, Beasley D B. Response of PM characteristics to NH<sub>3</sub> and other gaseous emissions at a southeaster layer operation. *ASABE Annual International Meeting*, 2009.
- [96] Walker J T, Whittall D R, Robarge W, Paerl H W. Ambient ammonia and ammonium aerosol across a region of variable ammonia emission density. *Atmospheric Environment*, 2004; 38(9): 1235–1246.
- [97] Hristov A N. Technical note: contribution of ammonia emitted from livestock to atmospheric fine particulate matter (PM 2.5) in the United States. *Journal of Dairy Science*, 2011; 94(6): 3130–3136.
- [98] USDA. Agricultural Air Quality Task Force White Paper. Ammonia emissions: what to know before you regulate. 2014. <http://www.soils.usda.gov/wps/portal/nrcs/detail/national/air/taskforce/?cid=stelprdb1268645>. Accessed on [2015-03-15].